

WASHING GLASSWARE(cont.)

PIPETS

To wash pipets: 30 min soapy water (dilute alconox)
 30 min (cold) tap water rinse
 15 min distilled tap rinse

Pipets should be picked up 2-3 times a day. I usually collect them in the morning, mid-afternoon, and around 5 p.m. **DON'T FORGET ABOUT TISSUE CULTURE PIPETS--** sometimes more can be collected here than in all the other buckets combined.

Before stuffing the pipets with cotton, check how many tissue culture vs. regular pipet cannisters are on the shelf--try to keep an even distribution of TC vs. regular. Make sure the cannister is appropriately labeled (ie. 25 TC vs. 25). Lately there seems to be frequent mix-ups. Plug the cotton neatly and firmly (but not too much--remember, we have to remove it before washing the pipets). And remember the trick of using 95% EtOH in a squirt bottle to easily remove the cotton plugs.

The cannisters get placed in the baking oven downstairs (the big blue one next to the ice machine). They should be put in before 6 p.m. (when the oven heat cycle starts), and placed spaced apart from one another. Do not set them in a stack or all in one spot because it will create a hot-spot and could possibly stop the cycle.

The pipets will be ready the next morning (as early as 6 a.m.) to put on the shelves. The autoclave tape will be brown from the oven, not striped like it is from the autoclave.

ALSO, the pipet buckets need to be changed once a week (sometimes more often--you'll know when the buckets are gooey or have changed color). Just rinse them out well with water and refill with water and a small amount of alconox. In addition, the tissue culture buckets get bleach added (approx. 20%).

YELLOW AND BLUE TIPS /EPPENDORF TUBES

We should have plenty of tips. The yellow ones are in small white boxes and the blue ones are in big boxes above the glassware racks or Lea's bench. Remember to put down clean bench paper and to wear gloves. Put a new piece of autoclave tape on each tip box (and remove any old pieces) and on the foil lid for the Eppendorf tube beaker.

Tips and tubes get **autoclaved** for: 30 min sterilize
 40 min dry
 GRAVITY CYCLE

Then put the tips and tubes in the drying oven (next to the sink) until they dry (usually at least 4 hours--overnight is okay).

TEST TUBES

All tubes get washed in the following procedure:

(wearing gloves--the orange ones above the sink are the best), squirt a small amount of soapy water into each tube, add water and scrub using an appropriate size brush (hanging above the sink). Rinse each tube 3 times with tap water and transfer to a new basket. Then rinse each tube again 3 times with distilled tap water. The caps to the culture tubes are washed in the same manner. Then, cap the culture tubes and put foil caps on the Corex tubes.

All glass tubes get put in a wire basket and put in the downstairs baking oven (the same one the pipets go in) for the overnight cycle.

PUT ONLY GLASS TUBES IN THE BAKING OVEN--ANY OTHER TUBES CAN SIMPLY BE DRIED IN THE DRYING OVEN NEXT TO THE SINK AND THEN PUT AWAY.

BOTTLED ddH₂O

Using clean/sterile bottles, fill to 500 ml with ddH₂O from the still.

Put bottles (with caps loosened) in autoclavable pan with small amount of water. Autoclave for 30 min. LIQUID CYCLE.

LB FOR MEDIA AND PLATES

For 1 liter:	Tryptone	10 g	NaCl	10g
	Yeast	5 g	ddH ₂ O to 1L	

For bottled media, dissolve into solution and aliquot into 500 ml bottles. Autoclave for 30 min.

For plates: Agar 20 g/liter

Autoclave for 30 min. LIQUID CYCLE

For amp. plates, once the media has cooled (until it is comfortable to hold using a paper towel), per liter add 500 µl of 100 mg/ml amp. stock.

Let the plates dry overnight at room temp. and put away in the cold room.

Don't forget to mark the **amp.** plates with a black marker--people do actually rely on that.

SPATULAS/STIR BARS

Wash with soapy water, rinse with water, then run under distilled tap water.
Dry and put away in the drawer.

Wash these twice a day--once in the morning and once in the late afternoon.

**AND IF YOU HAVE ANY QUESTIONS--DON'T HESITATE TO ASK ANYBODY
FOR HELP!**