

## IN-VITRO TRANSLATION

Prepare a master mix for n + 1 number of translations:

n = 1  
4.0  $\mu$ l sterile distilled water  
1.0  $\mu$ l 25X K-Mg  
2.0  $\mu$ l translation cocktail  
3.0  $\mu$ l  $^{35}\text{S}$ -met (5 mCi/ml)  
10.0  $\mu$ l nucleated rabbit reticulocyte lysate

20.0  $\mu$ l total per translation

Translation cocktail:

6.25 mM Spermidine HCl  
100 mM Creatine phosphate  
0.3125 mM amino acids (methionine minus)  
25 mM DTT  
250mMHEPES(pH7.4)

Add 20  $\mu$ l master mix to 5  $\mu$ l of a sample of (water + RNA). Typically, 0.5-1.0  $\mu$ g of polyA+ RNA or 5-20  $\mu$ g of total RNA is used for each 25  $\mu$ l translation. 5  $\mu$ l of water is used for the "endogenous" translation in the absence of RNA.

Incubate at 30°C for 90 minutes.

Note:

1. 25X K-Mg is a solution of potassium chloride and magnesium chloride (acetate salts can be used as well) such that 1X is the optimal concentration of K and Mg for the lysate. Typically K optimum is between 60-100 mM and Mg optimum can be from 0-0.1 mM.
2. Avoid repeated freeze/thaw of the methionine by distributing the stock into small aliquots. The chemical stability can be improved by diluting the stock to 5 mCi/ml with 1mM dithiothreitol. The amount of label can be increased to 35uCi or more per translation if so desired.

In order to determine the level of incorporation of  $^{35}\text{S}$ -met.:

1. Add a sample (2-5  $\mu\text{l}$ ) of the translation to 0.5 ml of water.
2. Add 0.5 ml of 1.0 N NaOH (to hydrolyze amino-acyl tRNA) and 80  $\mu\text{l}$  of 30% hydrogen peroxide (to decolorize).
3. Incubate at 37°C for 20 min.
4. Add 2.0 ml of 25% trichloroacetic acid containing carrier amino acids (2 mg/ml methionine or 3% casamino acids).
5. Chill on ice at least 20 min.
6. Collect the precipitate on glass fibre or nitrocellulose filters. Wash the filters with 10% TCA followed by 95% ethanol.
7. Allow the filters to dry. Count by liquid scintillation using an organic counting cocktail.
8. Typically, RNA should increase the incorporation of label 20-80 fold over endogenous. The conditions used here give 500-2000 cpm for endogenous translation and 30,000-100,000 cpm for samples with RNA in 5 $\mu\text{l}$ .
9. For analysis by SDS-PAGE, dilute up to 5  $\mu\text{l}$  of the translation assay with 20-30  $\mu\text{l}$  of sample buffer. Because of the large amount of protein in the lysate it is easy to exhaust the capacity of the SDS in the sample buffer; I therefore recommend using sample buffer that contains 4% SDS.